

## Controlled Release of bFGF from Elastomeric Biodegradable Microporous Sheets

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### Introduction

For wound healing applications in which the underlying traumatized tissue will experience substantial volumetric changes due to edema, it would be attractive to have a barrier dressing that could be sutured to the healthy wound periphery, but readily distend and contract to meet the mechanical demands imposed by the underlying tissue. Controlled release of a growth factor to facilitate wound healing from this material could provide added benefit. In an effort to meet such requirements we report here on the development of a thin, microporous elastic sheet with high tensile strength made from a biodegradable poly(ester urethane)urea (PEUU) loaded with the angiogenic basic fibroblast growth factor (bFGF) and processed with electrospinning. Characterization included assessing morphology, bFGF release and bioactivity, cell adhesiveness, and tensile properties.

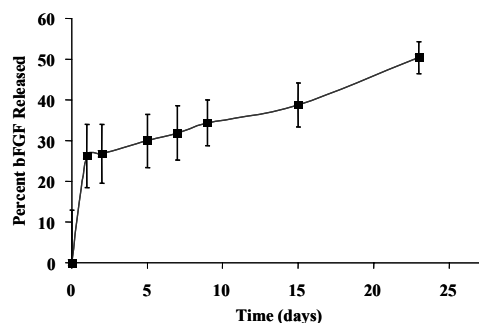
### Materials and Methods

PEUU was synthesized from polycaprolactone diol and 1,4-diisocyanatobutane with chain extension by putrescine as previously reported.<sup>1</sup> Bovine serum albumin (BSA) and bFGF (100:1) were dissolved in buffer and the solution was lyophilized. PEUU was dissolved at 6wt% in hexafluoroisopropanol and bFGF/BSA was added to the polymer solution at 500ng bFGF / mg PEUU. After solubilization the solution was electrospun over a 15-cm distance using a 1mL/min feed rate and by charging the solution at 10kV and the aluminum target at -10kV in a manner similar to that previously reported.<sup>2</sup>

Scanning electron microscopy (SEM) was utilized to characterize fiber morphologies in the sheets. Strips of material were placed in DMEM with 0.5% fetal bovine serum and 1% penicillin /streptomycin at 37°C for bFGF release studies. Immunoassay quantified bFGF concentration in the medium. Released bFGF bioactivity was measured by mitogenic assay. Briefly, rat vascular smooth muscle cells (SMCs) were seeded at  $1.5 \times 10^5$  in a 96-well plate. Media was replaced at 4h with the appropriate bFGF containing degradation media and cell number was measured 48 h later using the MTT mitochondrial activity assay. For cell adhesion, SMCs were seeded onto sheets at a density of  $2.0 \times 10^5$  cells/mL and cell adhesion was quantified 1 day after seeding using MTT. Tensile testing was completed according to ASTM D638-98.

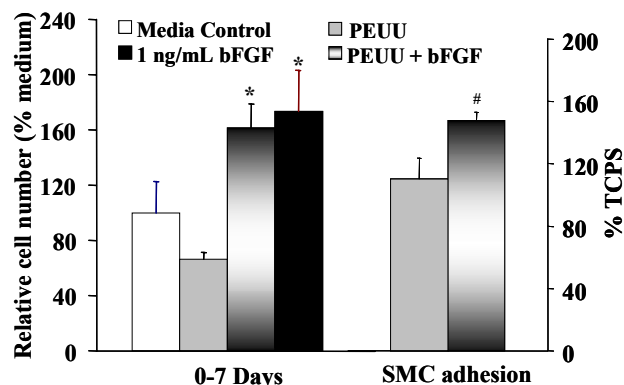
### Results

Electrospun sheets of PEUU and PEUU with bFGF consisted of continuous, bead-free sub-micron diameter fibers as observed by SEM. bFGF release from sheets consisted of an initial 25% burst followed by slower release up to 50% by 3 wks (**Figure 1**). Bioactivity



**Figure 1.** Release of bFGF from electrospun PEUU.

results from 0-7 days showed a statistically similar SMC mitogenic effect from PEUU loaded with bFGF compared to a control group directly receiving 1 ng/mL bFGF (**Figure 2**). Significantly higher SMC adhesion at  $148 \pm 5\%$  of tissue culture polystyrene (TCPS) was observed on PEUU loaded with bFGF relative to both TCPS and electrospun PEUU alone ( $p < 0.05$ ). No significant differences in tensile properties were found between PEUU and PEUU loaded with growth factor.



**Figure 2.** Released bFGF bioactivity at 1wk using a SMC mitogenic assay (left) and SMC adhesion on PEUU loaded with bFGF (\* $p < 0.05$  vs. medium control, # $p < 0.05$  greater than PEUU control).

### Conclusions

A biodegradable polyurethane was loaded with bFGF and processed into microporous sheets comprised of sub-micron scale fibers. These elastomeric sheets were capable of bioactive bFGF release for at least 1 week and facilitated increased SMC adhesion. In wound healing applications where material flexibility and growth factor release from a barrier would be desirable, these growth factor loaded sheets may provide attractive functionality.

### References

1. Guan J et al. *J Biomed Mater Res* 61:493 (2002).
2. Stankus JJ et al. *J Biomed Mater Res* 70:603 (2004).