

## Effects of a mixture of growth factors and serum proteins on human osteogenic cell cultures

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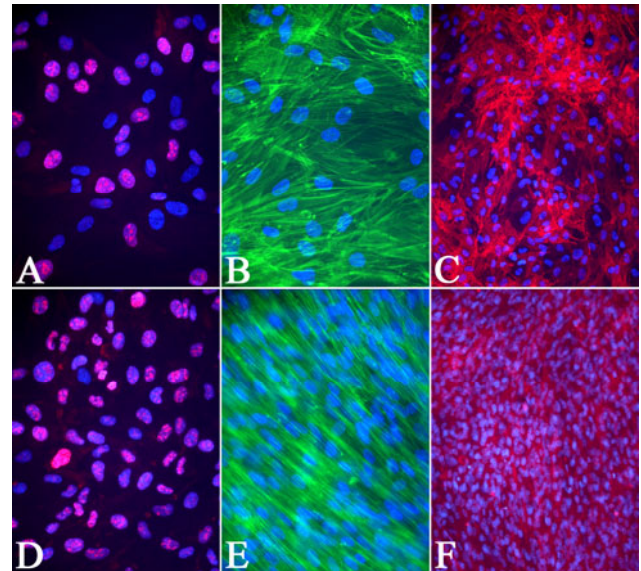
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**Introduction:** Platelet rich plasma (PRP) has been used in a series of clinical procedures to promote bone healing. Although beneficial clinical results have been reported, PRP has not been demonstrated to enhance bone formation under experimental conditions [1,2]. The aim of the present study was to evaluate the effects of a mixture of growth factors (GFs) and serum proteins that is typical of platelet extracts on various parameters of *in vitro* osteogenesis in human alveolar bone-derived cell cultures.

**Methods:** Osteoblastic cells were obtained by enzymatic digestion of human alveolar bone fragments and cultured under standard osteogenic condition until subconfluence. First passage cells were cultured ( $2 \times 10^4$  cells/well) on polystyrene (24-well plates and Thermanox coverslips) up to 14 days. The mixture of GFs and serum proteins tested contained 0.27  $\mu\text{g/ml}$  PDGF-BB, 0.22  $\mu\text{g/ml}$  TGF- $\beta$ 1, 0.15  $\mu\text{g/ml}$  TGF- $\beta$ 2, 37  $\mu\text{g/ml}$  albumin, 20  $\mu\text{g/ml}$  fibronectin, and 5  $\mu\text{g/ml}$  thrombospondin, all purchased from Sigma-Aldrich (St. Louis, MO). Cultures were exposed during the first 7 days (proliferative phase) to the mixture GFs+proteins in which PDGF-BB concentration was 20 ng/ml. For control cultures, the mixture was replaced by Gibco  $\alpha$ -MEM (Invitrogen, Carlsbad, CA). Cell morphology was observed by fluorescence labeling of actin cytoskeleton (Alexa Fluor-conjugated phalloidin; Molecular Probes, Eugene, OR) and nuclear stain (DAPI, Molecular Probes) at days 1, 4, 7, and 14; alkaline phosphatase (ALP) labeling was performed using a primary mouse anti-human ALP antibody (B4-78, Hybridoma Bank, Iowa City, IA) for cultures at day 7. Total cell number ( $\times 10^4$ ) and cell viability (% viable cells) were determined at days 1 and 4 by the Trypan blue exclusion assay. The proportion of cycling cells at days 1 and 4 was determined by double nuclear labeling DAPI/Ki-67 using a primary rabbit anti-human Ki-67 antibody (Diagnostic Biosystems, Pleasanton, CA). Alkaline phosphatase (ALP) activity normalized for total protein at day 4 ( $\times 10^{-7}$  mol p-nitrophenol/mg/min) and bone-like nodule formation (Alizarin red S (AR-S) staining) at day 14 were also evaluated. Data were compared by Mann-Whitney test.

**Results/Discussion:** At days 4, 7, and 14 epifluorescence clearly revealed that cultures exposed to the mixture of GFs and proteins exhibited a significantly higher number of adherent cells. In such experimental condition, the direction of the long axis of the cells did not significantly change throughout the culture mainly at days 7 and 14 (compare in Fig. 1, E with B). Although at day 1 there appeared to be less adherent cells in the control cultures, cell counts revealed no significant differences between groups ( $1.5 \pm 0.5$  for control and  $1.4 \pm 0.1$  for GFs+proteins;  $p > 0.05$ ). Total cell number at day 4 was  $5.8 \pm 0.8$  for the control cultures and  $25.2 \pm 5.6$  for the treated group ( $n=5$ ,  $p < 0.05$ ). At day 4, GFs+proteins cultures exhibited significantly higher values for total protein content ( $53.4 \pm 10.7$ ; for control,  $27.6 \pm 2.7$ ;  $n=5$ ,  $p < 0.05$ ) and reduced levels for ALP activity ( $0.25 \pm 0.2$ ; for control,  $3.9 \pm 0.25$ ;  $n=5$ ,  $p < 0.05$ ). Number of Ki-67 positive cells

(cycling cells) were significantly higher both at days 1 and 4 (Fig. 1A,D) for treated cultures compared to control ones (respectively at day 1,  $80 \pm 6.8$  and  $56.5 \pm 11.3$ ; at day 4,  $86.7 \pm 4.9$  and  $59.6 \pm 8.3$ ;  $n=3$ ,  $p=0.05$  for both comparisons). Cell viability was significantly higher for GFs+proteins exposed cultures compared to control ones ( $96.5 \pm 1.1$  and  $91 \pm 5.1$ , respectively;  $n=5$ ,  $p < 0.05$ ). Simple observation revealed more ALP positive cells with more intense labeling at day 7 (Fig. 1C,F) and more AR-S stained nodules at day 14 for control cultures.



**Figure 1.** Epifluorescence of osteogenic cells cultured under control conditions (A-C) and in the presence of GFs+proteins (D-F). Note that higher number of cells are observed for treated cultures at days 4 (A,D) and 7 (B,E;C,F). (A,D) Double nuclear labeling DAPI(blue)-Ki-67(red) shows higher number of Ki-67 positive cells in GFs+proteins cultures at day 4. (B,E) Double labeling phalloidin(green)-DAPI(blue) reveals that cells are oriented in the same direction in treated cultures at day 7. (C,F) ALP labeling (red) is clearly more intense for control cultures at day 7. Objectives: A,B,D,E = X40; C,F = X20.

**Conclusions:** The present results point toward a significant influence of a mixture of GFs and serum proteins on cell cycle during the proliferative phase of human osteogenic cell cultures, leading to an increase in the cell population and a reduction in the differentiation process. Strategies to improve bone healing should also include the presence of osteogenic factors such as BMPs during the onset of differentiation phase of cell cultures.

**References:** [1] Ranly DM, McMillan J, Keller T, Lohmann CH, Meunch T, Cochran DL, Schwartz Z, Boyan BD. J Bone Joint Surg Am 87:2052-2064, 2005. [2] Klongnoi B, Rupprecht S, Kessler P, Thorwarth M, Wiltfang J, Schlegel KA. Clin Oral Implants Res 17:312-320, 2006.

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